ClearBand Bradford Reagent

500 ml

Cat No: BR05

Shipping : Ship at ambient temperature. Storage : Stable for 1 year at 4°C.

General Information

ClearBand Bradford Reagent is formulated for a ready-to-use total protein analysis reagent used for quick measurement of total protein concentration. *ClearBand* Bradford Reagent contains Coomassie Brilliant Blue G-250, which associates with basic and aromatic amino acids, thus leading to a shift in absorbance during protein determination.

ClearBand Bradford Reagent offer an easy-to-use assay in either test tube or microplate format: mix protein sample with the assay reagent, incubate shortly and measure the absorbance at 595nm. Color response with Coomassie is non-linear with increasing protein concentration, therefore, a standard curve must be created with each assay.

Protocol (Standard Test Tube, Range: 25-1500 µg/ml)

1. Prepare a set of protein standards in the following concentrations with Bovine Serum Albumin (BSA, not provided) using the instructions provided in Table 1. As an initial step, dissolve 4 mg of BSA in 2 ml of ultrapure water. Refer this as Stock BSA that is in 2 mg/ml concentration. Use ultrapure water as diluent to prepare the standard set.

Table 1. Preparation	of BSA	Standards
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Standard	Diluent	BSA	Final Concentration
1	0	300 μL Stock	2000 μg/ml
2	150 μL	450 μL Stock	$1500 \ \mu g/ml$
3	325 μL	325 μL Stock	$1000 \ \mu g/ml$
4	325 μL	$325~\mu L$ Standard 2	$750 \ \mu g/ml$
5	325 μL	$325~\mu L$ Standard 3	$500 \mu g/ml$
6	325 μL	$325~\mu L$ Standard 5	$250~\mu g/ml$
7	325 μL	$325~\mu L$ Standard 6	$125 \mu g/ml$
8	$400~\mu L$	$100~\mu L$ Standard 7	25 μg/ml
9	$400\;\mu L$	0 μL	Blank

^{2.} Mix the *ClearBand* Bradford Reagent before use by gently inverting the bottle several times (Do not shake the bottle). Remove the amount of reagent needed and warm it to room temperature.

^{3.} Add 20 µl of each standard and unknown samples into appropriately labeled test tubes at least in triplicates.

^{4.} Add 1 ml of *ClearBand* Bradford Reagent to each tube and mix well.

- **5.** Incubate samples for 10 minutes at room temperature.
- **6.** Set the spectrophotometer to 595 nm. Zero the instrument on a cuvette filled only with water. Subsequently, measure the absorbance of all samples.
- 7. Subtract the average 595 nm measurement for the blank replicates from all other individual readings.
- 8. Prepare a standard curve by plotting the average blank-corrected 595nm measurement for each BSA standard vs. its concentration in $\mu g/mL$. Use the standard curve to determine the protein concentration of each unknown sample.

Protocol (Microplate, Range: 25-1500 µg/ml)

1. Prepare a set of protein standards in the following concentrations with Bovine Serum Albumin (BSA, not provided) using the instructions provided in Table 2. As an initial step, dissolve 4 mg of BSA in 2 ml of ultrapure water. Refer this as Stock BSA that is in 2 mg/ml concentration. Use ultrapure water as diluent to prepare the standard set.

Table 2. Preparation of BSA Standards

Standard	Diluent	BSA	Final Concentration
1	0	30 μL Stock	2000 μg/ml
2	15 μL	45 μL Stock	$1500 \ \mu g/ml$
3	32.5 μL	32.5 µL Stock	$1000~\mu g/ml$
4	32.5 μL	$32.5~\mu L$ Standard 2	750 μg/ml
5	32.5 μL	$32.5~\mu L$ Standard 3	$500 \mu g/ml$
6	32.5 μL	$32.5~\mu L$ Standard 5	250 μg/ml
7	32.5 μL	$32.5~\mu L$ Standard 6	125 μg/ml
8	40 μL	$10~\mu L$ Standard 7	25 μg/ml
9	$40~\mu L$	0 μL	Blank

- **2.** Mix the *ClearBand* Bradford Reagent before use by gently inverting the bottle several times (Do not shake the bottle). Remove the amount of reagent needed and warm it to room temperature.
- 3. Add 30 µl of each standard and unknown samples into appropriately labeled test tubes at least in triplicates.
- **4.** Add 250 µl of *ClearBand* Bradford Reagent to each tube and mix thoroughly on plate shaker for 30 seconds.
- **5.** Incubate samples for 10 minutes at room temperature.
- **6.** Set the spectrophotometer to 595 nm. Zero the instrument on a cuvette filled only with water. Subsequently, measure the absorbance of all samples.
- 7. Subtract the average 595 nm measurement for the blank replicates from all other individual readings.
- 8. Prepare a standard curve by plotting the average blank-corrected 595nm measurement for each BSA standard vs. its concentration in $\mu g/mL$. Use the standard curve to determine the protein concentration of each unknown sample.